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THREE NEW PHLEGMARIURINE B TYPE *LYCOPODIUM* ALKALOIDS FROM *HUPERZIA SERRATA*

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Phlegmariurine B (**1**), a known alkaloid, along with three new analogous compounds, 2 α -hydroxyphlegmariurine B (**2**), 2-oxophlegmariurine B (**3**) and 11-oxophlegmariurine B (**4**), were isolated from the CHCl₃ fraction of total alkaloids of whole plant of the Chinese medicinal herb *Huperzia serrata*. Their structures were elucidated by spectral analysis.

Keywords: *Huperzia serrata*; *Lycopodium* alkaloids; 2 α -Hydroxyphlegmariurine B; 2-Oxophlegmariurine B; 11-Oxophlegmariurine B

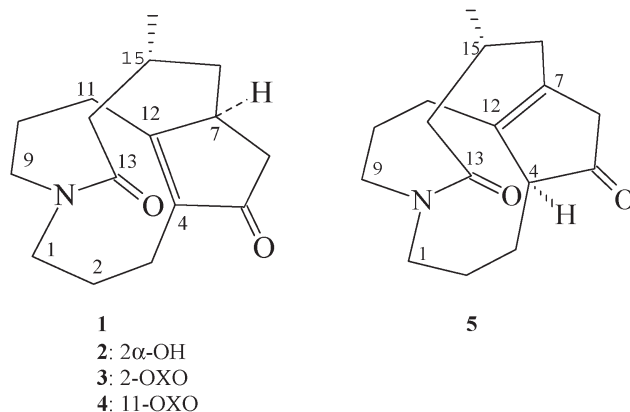
INTRODUCTION

Phlegmariurine B (**1**), isolated successively from *Phlegmariurus fordii* (Baker). Ching [1] and *Huperzia serrata* (Thunb.) Trev [2] of *Lycopodium* plant, was a unique structural type as *seco*-fawcettimine between C12 and C13 among *Lycopodium* alkaloids [3]. As a continuation of chemical constituents studies on *Huperzia serrata* [4–6], we reexamined the CHCl₃ extract of the total alkaloids of this plant, obtained **1** and three new analogues, 2 α -hydroxyphlegmariurine B (**2**), 2-oxophlegmariurine B (**3**) and 11-oxophlegmariurine B (**4**). This paper describes the structure elucidation of those compounds (Scheme 1).

RESULTS AND DISCUSSION

Compound **1** was identified as phlegmariurine B based on analyses of EIMS, IR, ¹H, ¹³C and DEPT NMR spectra [2]. Its relative configuration was defined as 7 α , 15 α *via* chemical conversion from phlegmariurine A (**5**) [1], whose configuration was determined by X-ray analysis [7]. The complete assignments of ¹H- and ¹³C-NMR signals (Tables I and II) were established by using HMQC, DEPT and ¹H-¹H COSY spectra.

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SCHEME 1 Structures of **1**–**5**.

The molecular formula $C_{16}H_{23}NO_3$ for compound **2** was deduced from its HR-EIMS spectrum. In the IR spectrum, a band at 3361.4 exhibited the presence of a hydroxyl group, the others resembled those of **1**. Comparison the ^{13}C -NMR data (Table II) with those of **1**, the most important difference was the presence of an oxygen-bearing methine signal at δ 60.72 and the absence of a methylene at δ 19.73 in the former, which suggested **2** to be 2-hydroxylated phlegmariurine B. The orientation of 2-OH was assigned as α by the analyses of the 1H -NMR data (Table I) combining with the study of molecular model. Its 1H -NMR spectrum displayed the proton coupling constants $J_{1\alpha,2}$ was 9.9 Hz. According to *Karplus* Equation ($J = 4.22 - 0.5 \cos\alpha + 4.2 \cos^2\alpha$), the dihedral angle between H-2 and H-1 α was more close to 180° rather than 0° or 60° . Furthermore, from the viewpoint of biogenesis, **2** was a possible derivative from **1**, and **1** was a cage structure, the hydroxyl group of **2** might be obtained only from the *exo* (α) orientation. Therefore, compound **2** was assigned as 2 α -hydroxyphlegmariurine B.

Compound **3**, $C_{16}H_{21}NO_3$ (HR-EIMS found: M^+ 275.1542, calcd: 275.1521), obtained as white amorphous powder, showed a positive effect on *Dragendorff's* reagent. Its ^{13}C -NMR spectrum (Table II) showed a carbonyl carbon signal at δ 204.59, and two methylene carbons signals at δ 58.58 and 38.27, the others were nearly overlapped with those of C4–C16 of **1**. The 1H -NMR spectrum (Table I) indicated two pairs of AB model protons at δ 3.18 and δ 5.14 (1-CH₂, $J = 15.7$ Hz), and δ 3.29 and δ 3.69 (3-CH₂, $J = 17.4$ Hz), respectively, which confirmed **3** to be 2-oxophlegmariurine B.

Compound **4** was isolated as colorless prisms. The molecular formula was established as $C_{16}H_{21}NO_3$ by the HR-EIMS spectrum. In the ^{13}C -NMR spectrum (Table II), a conjugated carbonyl carbon signal at δ 199.96 replaced the C-11 signal at δ 28.91 in **1**, which suggested **4** to be 11-oxophlegmariurine B. Additionally, in contrast to those of **1**, the differences of δ_{C-4} and δ_{C-12} and decrease of δ_{C-7} of **4** were consistent with the conjugated effect and γ -gauche effect from 11-carbonyl group, respectively. Hence, **4** was formulated as 11-oxophlegmariurine B.

EXPERIMENTAL SECTION

General Experimental Procedures

Melting points were determined on a *Fisher-Johns* melting point apparatus and are uncorrected. CD spectra were taken on a *Jasco J-715* spectropolarimeter in $CHCl_3$. IR spectra

TABLE I ¹H-NMR data of compounds 1–4 (400 MHz, CDCl₃)

| Position | 1 | 2 | 3 | 4 |
|----------------|----------------------------|----------------------------|----------------------------|----------------------------|
| 1 α | 2.81 ddd (14.1, 11.4, 4.7) | 2.66 dd (13.3, 9.9) | 3.18 d (15.7) | 2.90 ddd (14.2, 12.5, 4.6) |
| β | 4.05 dt (14.1, 2.5) | 4.07 dd (13.3, 5.6) | 5.14 d (15.7) | 4.19 dd (14.2, 4.7) |
| 2 α | 1.41 dtt (14.0, 4.7, 2.4) | – | – | 1.59 dtt (13.7, 6.0, 1.5) |
| β | 2.44* | 4.60 tt (9.9, 5.3) | – | 2.49* |
| 3 α | 2.44* | 2.68 dd (13.2, 9.8) | 3.29 d (17.4) | 2.82 td (14.2, 1.5) |
| β | 2.62* | 2.73 dd (13.2, 5.0) | 3.69 d (17.4) | 2.49* |
| 6 α | 2.35 dd (19.0, 7.1) | 2.37 dd (19.3, 7.1) | 2.45* | 2.49 dd (19.8, 6.9) |
| β | 2.20 dd (19.0, 2.2) | 2.20 dd (19.1, 1.9) | 2.45* | 2.28 dd (19.8, 2.5) |
| 7 | 2.75 br.s | 2.76 br.s | 2.88 br.s | 3.47 br.s |
| 8 <i>endo</i> | 1.76 dd (15.0, 4.6) | 1.77 dd (14.9, 4.6) | 1.93* | 1.67 ddd (14.9, 4.2, 1.5) |
| <i>exo</i> | 1.90 dd (15.1, 7.7) | 1.90 dd (15.0, 11.2) | 1.93* | 1.79 ddd (15.0, 11.2, 3.8) |
| 9 α | 3.18 dt (15.3, 3.7) | 3.27 dt (15.1, 3.0) | 3.46 dt (15.5, 4.5) | 3.31 ddd (15.4, 6.6, 1.9) |
| β | 3.87 ddd (15.2, 11.5, 3.7) | 3.89 ddd (15.5, 12.5, 3.0) | 4.07 ddd (15.2, 11.0, 4.0) | 4.05 ddd (15.3, 13.5, 3.3) |
| 10 α | 2.73* | 2.82 qt (13.6, 4.1) | 2.44* | 3.59 td (13.5, 6.6) |
| β | 1.92* | 1.91* | 2.11 dt (15.7, 4.6) | 2.82 ddd (13.8, 3.3, 2.0) |
| 11 α | 2.64* | 2.64 td (14.1, 3.7) | 2.78* | – |
| β | 2.73* | 2.74 ddd (14.0, 5.0, 3.4) | 2.78* | – |
| 14 <i>endo</i> | 1.86 d (14.8) | 1.84 d (14.7) | 1.94 d (13.8) | 1.86 d (15.2) |
| <i>exo</i> | 2.43 dd (14.6, 8.5) | 2.40 dd (14.7, 8.7) | 2.46* | 1.97 dd (15.2, 7.8) |
| 15 <i>endo</i> | 2.12 m | 2.11 m | 2.46 m | 2.10 m |
| 16 | 1.07 d (7.0) | 1.05 d (7.0) | 1.09 d (6.4) | 1.03 d (7.0) |

* Overlapping signals.

TABLE II ^{13}C -NMR data of compounds **1**–**4** (100 MHz, CDCl_3)

| Position | 1 | 2 | 3 | 4 |
|----------|----------|----------|----------|----------|
| 1 | 51.12 t | 56.52 t | 58.58 t | 51.73 t |
| 2 | 19.73 t | 60.72 d | 204.59 s | 19.68 t |
| 3 | 23.36 t | 32.36 t | 38.27 t | 24.05 t |
| 4 | 141.69 s | 138.78 s | 136.06 s | 149.73 s |
| 5 | 207.35 s | 207.59 s | 205.76 s | 208.30 s |
| 6 | 38.49 t | 38.27 t | 37.67 t | 38.18 t |
| 7 | 41.15 d | 41.32 d | 41.54 d | 36.20 d |
| 8 | 40.95 t | 40.86 t | 39.98 t | 40.32 t |
| 9 | 50.98 t | 51.50 t | 49.76 t | 48.26 t |
| 10 | 25.77 t | 25.49 t | 26.10 t | 44.69 t |
| 11 | 28.91 t | 28.81 t | 28.27 t | 199.96 s |
| 12 | 171.65 s | 174.22 s | 174.38 s | 162.63 s |
| 13 | 173.30 s | 173.94 s | 175.66 s | 173.91 s |
| 14 | 40.75 t | 40.30 t | 39.09 t | 40.71 t |
| 15 | 27.11 d | 27.24 d | 27.83 d | 26.44 d |
| 16 | 26.59 q | 26.42 q | 25.41 q | 26.09 q |

were recorded on a *Nicolet Magna 750 FTIR* (KBr) spectrophotometer. EIMS and HREIMS data were obtained with *MAT-95* and *MAT-711* mass spectrometer. NMR spectra were recorded on a *Bruker AM-400* instrument with CD_3Cl as solution and residual CHCl_3 peak (δ_{H} : 7.26; δ_{C} : 77.10) as reference with chemical shift δ in 1 and *J* in Hz. Optical rotations were measured using a *P.E. 241 MC* polarimeter in CHCl_3 . Silica gel (200–300, 400 mesh) was used for column chromatography (CC) and precoated plates of silica gel (HSGF₂₅₄) for TLC.

Plant Material

Fresh whole plant of *Huperzia serrata* (Thunb) Trev. (Huperziaceae) was collected in Zhejiang Province of China and identified by Dr Xiao-Qiang Ma. Voucher sample (No. 97-63) was deposited in the Herbarium of this institute.

Extraction and Isolation

The air-dried whole plants (10 kg) were powdered and extracted with 1% aqueous tartaric acid at room temperature and the concentrated acidic extract basified to pH9 with concentrated NH_3 solvents, then extracted with CHCl_3 . The CHCl_3 layer was concentrated to obtain the CHCl_3 fraction of total alkaloid extract, which was chromatographed over silica gel (1 kg) column with gradient eluants (CHCl_3 , 1000 ml; 1–4% methanol in CHCl_3 , each 1000 ml) to afford fr. 1–5. Fr. 3 (2.0 g) was chromatographed on silica gel column eluting with EtOAc/acetone (2:1, 1200 ml), collected every 100 ml volume, gained 3 fr.s: fr. 3.1–3.4, fr. 3.5–3.7, and fr. 3.8–3.12. Vaporized solvent under reduced pressure, and dissolved with 10 ml acetone, respectively. Fr. 3.5–3.7 yielded crude crystals, recrystallized twice with acetone resulting in phlegmariurine B (**1**, 750 mg); the concentrated mother liquid (235 mg) was subjected to silica gel (20 g) CC with $\text{CHCl}_3/\text{CH}_3\text{OH}$ (20:1, 480 ml), collected with 10 ml tubes and detected using TLC (silica gel HSGF₂₅₄, $\text{CHCl}_3/\text{CH}_3\text{OH}$ 15:1, iodine vapor for detection); yielding **2** (13 mg, R_f : 0.42), **3** (3 mg, R_f : 0.51), and **4** (15 mg, R_f : 0.62).

Phlegmariurine B (**1**): Colorless prisms from acetone, 750 mg, mp 110–111°C; CD (c 3.9×10^{-4} , CHCl_3): $[\theta]_{255} - 5.8 \times 10^4$; IR (KBr) ν_{max} cm^{-1} : 3363.3, 2921.7, 1687.4,

1643.2, 1623.8, 1457.9, 1417.4, 1234.2, 1091.5, 817.7; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) see Tables I and II; EIMS (m/z): 261 (M^+ , 100), 246 (20), 233 (23), 218 (22), 190 (30), 176 (30), 150 (55).

2 α -Hydroxyphlegmariurine B (2): Colorless prisms from acetone, 13 mg, mp 240–241°C; $[\alpha]_{\text{D}}^{25}$: -0.17 (c 0.17, CHCl_3); CD (c 4.3×10^{-4} , CHCl_3): $[\theta]_{226} - 4.4 \times 10^4$; IR (KBr) $\nu_{\text{max}} \text{cm}^{-1}$: 3361.4, 2915.9, 1691.3, 1641.2, 1604.5, 1465.7, 1039.5; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) see Tables I and II; EIMS (m/z): 277 (M^+ , 65), 258 (8), 259 (17), 249 (94), 234 (100), 220 (35), 206 (39), 192 (30), 178 (42), 164 (37), 150 (28); HR-EIMS: 277.1687 (M^+), calcd for $\text{C}_{16}\text{H}_{23}\text{NO}_3$, 277.1678.

2-Oxo-phlegmariurine B (3): White amorphous powder, 3 mg, $[\alpha]_{\text{D}}^{25}$: -2.22 (c 0.033, CHCl_3); CD (c 2.2×10^{-4} , CHCl_3): $[\theta]_{231} + 1.6 \times 10^5$, $[\theta]_{259} - 1.3 \times 10^5$; IR (KBr) $\nu_{\text{max}} \text{cm}^{-1}$: 3434.7, 2925.5, 2850.3, 1731.8, 1668.2, 1456.0, 1053.0; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) see Tables I and II; EIMS (m/z): 275 (M^+ , 64), 248 (9), 232 (9), 219 (16), 204 (14), 192 (14), 183 (18), 176 (21), 170 (24), 154 (78), 150 (42), 123 (22); HR-EIMS: 275.1542 (M^+), calcd for $\text{C}_{16}\text{H}_{21}\text{NO}_3$, 275.1521.

11-Oxo-phlegmariurine B (4): Colorless prisms from acetone-petroleum ester, 15 mg, mp 233–235°C; $[\alpha]_{\text{D}}^{25}$: -5.08 (c 0.35, CHCl_3); CD (c 3.7×10^{-4} , CHCl_3): $[\theta]_{250} + 8.2 \times 10^4$, $[\theta]_{291} - 8.9 \times 10^4$; IR (KBr) $\nu_{\text{max}} \text{cm}^{-1}$: 3382.6, 2950.6, 2912.0, 1702.9, 1664.3, 1623.8, 1469.5, 1432.9, 1375.0, 1207.2, 1180.2, 1151.3, 1085.7, 954.6; $^1\text{H-NMR}$ (400 MHz, CDCl_3) and $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) see Tables I and II; EIMS (m/z): 275 (M^+ , 33), 247 (4), 232 (4), 220 (18), 219 (100), 163 (88), 154 (22), 148 (22), 121 (20); HR-EIMS: 275.1520 (M^+), calcd for $\text{C}_{16}\text{H}_{21}\text{NO}_3$, 275.1521.

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